



BAGP POLE NGS testing guidance, v1.1, dated 8 April 2022

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BAGP guidance on *POLE* NGS testing in endometrial carcinoma

GLOSSARY:

EC: Endometrial carcinoma

G1/G2/G3/low-grade/high-grade: FIGO grades of endometrioid carcinoma; note that this grading system does not apply to non-endometrioid histotypes which are broadly classified as 'high-grade'

Histotype: Histological type of EC categorised on morphological features as defined by the WHO

IHC: Immunohistochemistry

LVSI: Lymphovascular space invasion

MMR: Mismatch repair

NGS: Next Generation Sequencing

NSMP: No Specific Molecular Profile, i.e. EC that do not show MMR, p53 or *POLE* defects

POLE: DNA polymerase epsilon

Stage: FIGO or TNM staging of EC

WHO: World Health Organisation (specifically the classification of female genital tumours, 5th edition, 2020)

Note 1: *POLE* NGS testing in endometrial carcinoma

Molecular testing in endometrial carcinoma, including mutational analysis for *POLE*, is recommended by the World Health Organisation wherever resources permit¹. *POLE* NGS testing will soon be available for NHS patients via national genomics services within the UK. This requires considerable resources and additional work for laboratories in sample preparation and transport, sample analysis and provision of an integrated report. This algorithm is provided to limit *POLE* testing to those cases where it is essential for patient care. This is meant to be a guideline and it is appreciated that centres may follow local policy on the extent of testing and opt to further minimise and restrict testing to only those cases where the *POLE* result alters decisions regarding adjuvant treatment, or at the other extreme, move to universal testing of all cases. Interpretation of *POLE* results should be based on published data²: 11 mutations have been established as being pathogenic, with the variants in bold accounting for the majority of cases: **P286R (c.857C>G)**, **V411L (c.1231G>T/C)**, **S297F (c.890C>T)**,

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S459F (c.1376C>T), A456P (c.1366G>C), F367S (c.1100T>C) L424I (c.1270C>A), M295R (c.884T>G), P436R (c.1307C>G), M444K (c.1331T>A) and D368Y (c.1102G>T).

Non-pathogenic variants and guidance for determining the pathogenicity of additional variants has also been put forward².

Note 2: Testing of all endometrial carcinoma biopsies regardless of histotype

Following the discovery of the four molecular subtypes of endometrial cancer (EC) by The Cancer Genome Atlas (TCGA) in 2013³, the **Proactive Molecular Risk Classifier for Endometrial Cancer (ProMisE)** was developed and validated for clinical use⁴⁻⁶. This uses a combination of targeted sequencing for pathogenic *POLE* exonuclease domain mutations and immunohistochemistry (IHC) to determine MMR and p53 status, to assign EC to one of four molecular subtypes; *POLE*mut, MMRd, p53abn, and NSMP (no specific molecular profile). Molecular classification of EC has since been shown to have a stronger prognostic significance than conventional histological classification⁴⁻⁸. Historically, the EC risk stratification used to guide treatment has largely been based on histological type, grade, and stage. Both histotype and grade assignment have been shown to be poorly reproducible, even amongst expert pathologists, with no agreement in histotype diagnosis in up to one third of cases⁹⁻¹¹. In contrast, EC molecular classification has been shown to be highly reproducible and can be accurately performed on endometrial biopsies, with high concordance between biopsy and final hysterectomy specimen^{12,13}. The 2020 5th edition of the WHO Female Genital Tumours recommends the integration of molecular classification into standard EC pathology reporting¹. While the assessment of MMR and p53 IHC are widely available, cost and access to *POLE* testing currently presents a major barrier to widespread adoption. This algorithm presents a way of restricted use of *POLE* testing to patients whom *POLE* status would impact on treatment decisions.

Note 3: MMR IHC should be performed on all endometrial cancers

As recommended by NICE in October 2020 (Diagnostics guidance DG42 available at: [Overview | Testing strategies for Lynch syndrome in people with endometrial cancer | Guidance | NICE](#)), all patients diagnosed with EC should have screening for Lynch syndrome¹⁴. The initial step is four-panel IHC testing for MMR deficiency (MLH1, MSH2, MSH6 and PMS2). If the IHC is abnormal, with loss of MLH1 +/- PMS2, MLH1 promoter hypermethylation testing on tumour DNA should be performed to differentiate sporadic and Lynch syndrome-associated cancers. If loss of MSH2, MSH6, or isolated PMS2 is found on IHC, germline testing for Lynch syndrome is recommended¹⁴. Identification of mismatch repair deficient (MMRd) EC also has important therapeutic implications. Defects in mismatch repair lead to a high mutational burden, an abundance of neoantigens and profound immune response, making these ECs an ideal target for immune checkpoint blockade (ICB) therapy. The 2017 FDA approval for ICB therapy in MMRd solid tumours has been followed by multiple single agent EC clinical trials with PD-1 and PD-L1 inhibitors showing response rates of 49-57% and 27-43%, respectively, in MMRd advanced, metastatic or recurrent ECs¹⁵.

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Note 4: p53 IHC should be performed on all endometrial cancers

p53abn EC represent the most aggressive molecular subtype, and account for majority of EC mortality¹⁶. The ECs in this molecular subtype were characterised by TCGA as having a very high number of somatic copy number alterations, low mutation rate, and ubiquitous *TP53* mutations³. These tumours are now identified by a more pragmatic method of mutant-pattern p53 IHC staining, which has been shown to be an excellent surrogate maker for *TP53* mutational status as determined by sequencing in EC biopsies¹⁷. Recent evidence has shown the p53abn molecular subtype is found in up to 5-10% of grade 1 and 2 endometrioid ECs^{16,18-20}. The 2020 ESGO/ESTRO/ESP guidelines classify any stage p53abn EC with myoinvasion, regardless of the grade or histotype, as high-risk disease, with a recommendation for adjuvant chemotherapy +/- radiation²¹. The value of carrying out universal p53 IHC is to avoid missing cases of serous carcinoma that may mimic endometrioid carcinoma, as well as identifying the infrequent p53abn EC cases that show endometrioid grade 1 or grade 2 histology.

Note 5: ER IHC should be performed on all endometrial cancers

The category of NSMP EC is one of exclusion, namely all those EC that do not harbour mismatch repair defects or pathogenic *POLE* or *TP53* mutations. This is a large and heterogeneous group that accounts for about 50% of all EC. Studies on biomarkers of poor outcome in this category have thus far been inconclusive, however, ER status and presence of LVSI. The value of universal ER testing is to indicate the possibility rarer histotypes of EC that may closely mimic endometrioid EC, such as clear cell, mesonephric-like and gastrointestinal-type mucinous EC; these are negative for ER, in contrast to the generally strong positivity typical of endometrioid EC. In addition, knowledge of ER status may help to inform decisions on hormonal treatment including hormone replacement therapy. Most importantly, ER status has been demonstrated to correlate with recurrence-free survival in NSMP EC²²; notably in this analysis a cut-off of 10% was applied and thereby flat negative or very focal ER expression should alert the pathologist to the possibility of a non-endometrioid histotype or potential for aggressive behaviour in an endometrioid tumour.

Note 6: Interpretation of MMR IHC

MMR status should be reported as normal or abnormal, with an appropriate message relating to the likelihood of Lynch Syndrome and thereby recommendation for referral to clinical genetics services. MMR IHC interpretation and reporting terminology have been the subject of previous BAGP guidance²³.

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Note 7: Interpretation of p53 IHC

IHC results of p53 should be reported as normal or abnormal (aberrant or mutation-type are appropriate alternatives). Reporting as 'positive' or 'negative' should be avoided. Pathologists should be familiar with the abnormal staining patterns: over-expression, complete absence (also known as 'null'), cytoplasmic and subclonal, as distinct from normal or 'wild-type' p53 expression. This has been the subject of previous BAGP guidance²⁴, an online teaching resource for p53 interpretation in EC ([Interpretation of p53 Immunostaining in Endometrial Carcinoma \(ubc.ca\)](#)) and detailed reviews^{25,26}.

Note 8: *POLE* testing in the presence of abnormal MMR and/or p53 IHC

Given the high mutational burden observed in *POLE*mut and MMRd tumours, secondary *TP53* mutations/p53 IHC abnormalities can be found. Leon-Castillo *et al* reported on the approximately 3-5% of ECs that have more than one molecular feature, called 'multiple classifiers'²⁷. They demonstrated that MMRd-p53abn tumours have morphology, molecular profiles and clinical behaviour aligning with MMRd EC, and the *POLE*mut-p53abn EC have morphology, molecular profiles and clinical behaviour aligning with *POLE*mut EC. These findings suggest the *TP53* mutation is a later event during tumour progression in *POLE*mut and MMRd tumours and does not affect the clinical outcome²⁷. This also highlights the importance of interpreting p53 and MMR IHC in the context of *POLE* mutation status to avoid overtreatment in these patients with 'multiple classifiers'. The 2020 ESGO/ESTRO/ESP guidelines for the management of patients with EC classify stage I-II *POLE*mut EC as low-risk and omission of adjuvant therapy should be considered²¹.

Note 9: Surgical staging including hysterectomy

FIGO staging for EC is surgical²⁸. Standard surgery consists of a total hysterectomy and bilateral salpingo-oophorectomy, and there is high level evidence supporting a minimally invasive surgical approach^{29,30}. Sentinel lymph node biopsy has been shown to have a high diagnostic accuracy in EC, and is now recommended to be performed over full lymphadenectomy¹⁴. Indications for nodal assessment in EC varies significantly between centres. The British Gynaecological Cancer Society (BGSC) recommends that surgical staging, including sentinel lymph node biopsy and omental biopsy, may be appropriate for women with disease greater than low risk¹⁴. For patients with known stage II EC, total hysterectomy and bilateral salpingo-oophorectomy is still recommended, and radical hysterectomy should only be performed if required to obtain tumour free margins^{14,21}. Further in-depth discussion regarding surgical staging in EC is beyond the scope of this document.

Note 10: Lymphovascular space invasion (LVSI) in EC

Lymphovascular space invasion (LVSI) is an established prognostic indicator in endometrial carcinoma and influences ESGO-ESTRO-ESP risk categorisation²¹. A binary distinction between no or focal LVSI as opposed to extensive/substantial LVSI is recommended but the cut-off has varied: previous recommendations by the International Society of Gynecological Pathologists defined 'extensive' LVSI

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as the presence of three or more vessels containing tumour³¹, while the 2020 WHO Classification and ESGO-ESTRO-ESP guidelines state the presence 5 or more involved vessels being definitional for extensive/substantial LVSI^{1,21}. The International Collaboration on Cancer Reporting dataset recommends: "Although there have been different proposals for what constitutes extensive LVI, it is a good rule of thumb to diagnose extensive LVI when it is easily recognisable at scanning magnification (and artefact is excluded) and when present in three or more vessels on closer inspection" .³²

Note 11: No *POLE* testing is indicated for low grade (grade 1 or 2) endometrioid, stage IA, no LVSI, and normal MMR and p53 IHC

Talhok *et al* recently validated an amended ProMisE testing protocol where p53 and MMR IHC were performed on all cases with omission of *POLE* sequencing in 'very low-risk' EC³³. These were defined as cases fulfilling all of the following: grade1/2, MMR-proficient, p53 wild-type, endometrioid histology, stage IA, no LVSI. The rationale for omitting *POLE* testing in this 'very low-risk' subgroup is because the clinical outcomes are excellent, with no adjuvant therapy required, thus *POLE* status would not alter management of these patients. In this validation series they did confirm this 'very low-risk' EC cohort with unknown *POLE* mutational status had excellent clinical outcomes, comparable to that of ECs known to have pathogenic mutations in *POLE*.

Note 12: *POLE* testing is recommended in Stage I-II non-endometrioid EC or any grade endometrioid stage IA with LVSI or stage IB /stage II regardless of LVSI

The majority of *POLE*mut ECs are endometrioid, and despite many having high-risk features, such as high-grade and LVSI, this molecular subtype has exceptionally favourable survival outcomes⁴⁻⁸. A recent meta-analysis which included 294 patients with pathogenic *POLE* mutations showed that the excellent survival outcomes of stage I/II *POLE*mut EC was independent of adjuvant treatment received³⁴. There are two clinical trials currently assessing the safety of de-escalation of adjuvant treatment in *POLE*mut EC (PORTEC-4a and TAPER- Tailored adjuvant therapy in *POLE*-mutated and p53-wildtype early stage endometrial cancer) and a third will be commencing shortly (the TransPORTEC RAINBO Blue-*POLE* arm). The 2020 ESGO/ESTRO/ESP guidelines classify stage I-II *POLE*mut EC as low risk and state that in women with Stage I-II pathogenic *POLE*mut ECs, omission of adjuvant therapy should be considered²¹. We therefore recommend testing *POLE* status in patients where the recommended adjuvant therapy would change if a pathogenic *POLE* mutation is found:

- Stage I-II non-endometrioid ECs
- Any grade endometrioid stage IA with LVSI, grade 3 endometrioid stage IA with no/focal LVSI, or any grade endometrioid stage IB/stage II regardless of LVSI

Identification of pathogenic *POLE* mutations in these patients will allow discussion of de-escalation of adjuvant therapy and/or identify patients eligible for de-escalation clinical trials and these patients may be spared the toxicity and cost of unnecessary radiation treatment +/- chemotherapy.

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Note 13: *POLE* testing is not indicated in Stage III-IV EC unless directed by MDT and/or patient choice

There is currently insufficient evidence to guide adjuvant treatment of advanced stage (III-IV) *POLE*mut EC. Furthermore, the 2020 EGSO/ESTRO/ESP EC treatment recommendations for stage III-IV EC are the same regardless of molecular subtype²¹, therefore *POLE* status would not currently alter management in these patients.

References

1. WHO Classification of Tumours. Female genital tumours: International Agency for Research on Cancer Lyon (France): International Agency for Research on Cancer; 2020.
2. León-Castillo A, Britton H, McConechy MK, McAlpine JN, Nout R, Kommos S, Brucker SY, Carlson JW, Epstein E, Rau TT, Bosse T, Church DN, Gilks CB. Interpretation of somatic *POLE* mutations in endometrial carcinoma. *J Pathol*. 2020 Mar;250(3):323-335.
3. Getz G, Gabriel SB, Cibulskis K, et al. Integrated genomic characterization of endometrial carcinoma. *Nature* 2013; 497: 67–73.
4. Talhouk A, McConechy MK, Leung S, et al. A clinically applicable molecular-based classification for endometrial cancers. *Br J Cancer* 2015; 113: 299–310.
5. Talhouk A, McConechy MK, Leung S, et al. Confirmation of ProMisE: A simple, genomics-based clinical classifier for endometrial cancer. *Cancer* 2017; 123: 802–813.
6. Kommos S, McConechy MK, Kommos F, et al. Final validation of the ProMisE molecular classifier for endometrial carcinoma in a large population-based case series. *Ann Oncol* 2018; 29: 1180–1188.
7. Stelloo E, Bosse T, Nout RA, et al. Refining prognosis and identifying targetable pathways for high-risk endometrial cancer; A TransPORTEC initiative. *Mod Pathol* 2015; 28: 836–844.
8. Stelloo E, Nout RA, Osse EM, et al. Improved risk assessment by integrating molecular and clinicopathological factors in early-stage endometrial cancer-combined analysis of the PORTEC cohorts. *Clin Cancer Res* 2016; 22: 4215–4224.
9. Gilks CB, Oliva E, Soslow RA. Poor interobserver reproducibility in the diagnosis of high-grade endometrial carcinoma. *Am J Surg Pathol* 2013; 37: 874–881.
10. Thomas S, Hussein Y, Bandyopadhyay S, et al. Interobserver variability in the diagnosis of uterine high-grade endometrioid carcinoma. *Arch Pathol Lab Med* 2016; 140: 836–843.
11. de Boer SM, Wortman BG, Bosse T, et al. Clinical consequences of upfront pathology review in the randomised PORTEC-3 trial for high-risk endometrial cancer. *Ann Oncol* 2018; 29: 424–430.
12. Talhouk A, Hoang LN, McConechy MK, Nakonechny Q, Leo J, Cheng A, Leung S, Yang W, Lum A, Köbel M, Lee CH, Soslow RA, Huntsman DG, Gilks CB, McAlpine JN. Molecular classification of endometrial carcinoma on diagnostic specimens is highly concordant with final hysterectomy: Earlier prognostic information to guide treatment. *Gynecol Oncol*. 2016 Oct;143(1):46-53.
13. Stelloo E, Nout RA, Naves LCLM, et al. High concordance of molecular tumor alterations between pre-operative curettage and hysterectomy specimens in patients with endometrial carcinoma. *Gynecol Oncol* 2014; 133: 197–204.
14. Morrison J, Balega J, Buckley L, et al. British Gynaecological Cancer Society (BGCS) uterine cancer guidelines: Recommendations for practice. *Eur J Obstet Gynecol Reprod Biol* 2022; 270: 50–89.
15. Green AK, Feinberg J, Makker V. GYNECOLOGIC CANCER A Review of Immune Checkpoint Blockade Therapy in Endometrial Cancer.
16. Jamieson A, Thompson EF, Huvila J, et al. p53abn Endometrial Cancer: understanding the most aggressive endometrial cancers in the era of molecular classification. *Int J Gynecol Cancer* 2021; ijgc-2020-002256.
17. Singh N, Piskorz AM, Bosse T, et al. p53 immunohistochemistry is an accurate surrogate for TP53

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- mutational analysis in endometrial carcinoma biopsies. *J Pathol* 2020; 250: 336–345.
18. Yano M, Ito K, Yabuno A, et al. Impact of TP53 immunohistochemistry on the histological grading system for endometrial endometrioid carcinoma. *Mod Pathol* 2019; 32: 1023–1031.
 19. Thompson E, Huvila J, Leung S et al. Refining pathologic interpretation of endometrial carcinomas: lessons learned from a nationwide study in a new era of molecular classification. *Int J Gynecol Cancer* 2020; A3-A4 (abstract only).
 20. Jamieson A, Thompson EF, Huvila J, et al. Endometrial carcinoma molecular subtype correlates with the presence of lymph node metastases. *Gynecol Oncol* 2022; 1–9.
 21. Concin AN, Matias-guiu X, Vergote I, et al. ESGO / ESTRO / ESP Guidelines for the management of patients with endometrial carcinoma. *Int J Gynecol Cancer* 2020; 1–63.
 22. Vermij L, Powell M, Leon-Castillo A, et al. Molecular profiling of NSMP high-risk endometrial cancers of the PORTEC-3 trial – prognostic refinement and druggable targets. *International Journal of Gynecol Cancer* 2021;31:A89-A90 (abstract only).
 23. Singh N, Wong R, Tchrakian N, Allen S-G, Clarke B, Gilks CB. Interpretation and Reporting Terminology for Mismatch Repair Protein Immunohistochemistry in Endometrial Cancer. [1593411202wpdm BAGP MMR IHC Interpretation June 2020.pdf \(thebagp.org\)](#). Accessed March 30, 2022.
 24. Kobel M, McCluggage WG, Gilks CB, Singh N. Interpretation of p53 Immunohistochemistry In Tubo-Ovarian Carcinoma: Guidelines for Reporting. <https://www.thebagp.org/download/bagp-ukneqas-project-p53-interpretation-guide-2016/>
 25. Köbel M, Ronnett BM, Singh N, Soslow RA, Gilks CB, McCluggage WG. Interpretation of p53 Immunohistochemistry in Endometrial Carcinomas: Toward Increased Reproducibility. *Int J Gynecol Pathol*. 2019 Jan;38 Suppl 1(Iss 1 Suppl 1):S123-S131.
 26. Wong RW-C, Palicelli A, Hoang L, Singh N. Interpretation of p16, p53 and mismatch repair protein immunohistochemistry in gynaecological neoplasia. *Diagnostic Histopathol* 2020; 26: 257-277.
 27. León-Castillo A, Gilvazquez E, Nout R, et al. Clinicopathological and molecular characterisation of ‘multiple-classifier’ endometrial carcinomas. *J Pathol* 2020; 250: 312–322.
 28. Amant F, Mirza MR, Koskas M, et al. Cancer of the corpus uteri. *Int J Gynecol Obstet* 2018; 143: 37–50.
 29. Janda M, Gebiski V, Davies LC, et al. Effect of total laparoscopic hysterectomy vs total abdominal hysterectomy on disease-free survival among women with stage i endometrial cancer: A randomized clinical trial. *JAMA - J Am Med Assoc* 2017; 317: 1224–1233.
 30. Walker JL, Piedmonte MR, Spirtos NM, et al. Recurrence and survival after random assignment to laparoscopy versus laparotomy for comprehensive surgical staging of uterine cancer: Gynecologic Oncology Group LAP2 study. *J Clin Oncol* 2012; 30: 695–700.
 31. Singh N, Hirschowitz L, Zaino R, Alvarado-Cabrero I, Duggan MA, Ali-Fehmi R, Euscher E, Hecht JL, Horn LC, Ioffe O, Matias-Guiu X, McCluggage WG, Mikami Y, Ordi J, Parkash V, Qudus MR, Quick CM, Staebler A, Zaloudek C, Nucci M, Malpica A, Oliva E. Pathologic Prognostic Factors in Endometrial Carcinoma (Other Than Tumor Type and Grade). *Int J Gynecol Pathol*. 2019 Jan;38 Suppl 1(Iss 1 Suppl 1):S93-S113.
 32. International Collaboration on Cancer Reporting Dataset for Endometrial Cancers. [Endometrial Cancers - International Collaboration on Cancer Reporting \(iccr-cancer.org\)](#)
 33. Talhouk A, Jamieson A, Crosbie EJ, Taylor A, Chiu D, Leung S, Grube M, Kommos S, Gilks CB, McAlpine JN, Singh N. Targeted Molecular Testing in Endometrial Carcinoma: Validation of a Clinically Driven Selective ProMisE Testing Protocol. *Int J Gynecol Pathol*, 2022, under review.
 34. McAlpine JN, Chiu DS, Nout RA, et al. Evaluation of treatment effects in patients with endometrial cancer and POLE mutations: An individual patient data meta-analysis. *Cancer* 2021; 1–14.

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